



# Test Training Xpert® GI Panel

For use with:  
GeneXpert® Dx System  
GeneXpert® System with Touchscreen  
GeneXpert® Infinity System

*Catalog Number : GXGI-CE-10*

*For CE-IVD Only*

*CE-marked in accordance with IVDR (Regulation (EU) 2017/746)*

CE<sup>2797</sup> IVD In Vitro Diagnostic Medical Device



303-7273 Rev. A April 2026



# Training Objectives

*At the end of the training, users will be able to:*

- Properly store and handle the Xpert® GI Panel cartridge kit and specimen collection
- Follow proper laboratory safety precautions
- Collect and transport appropriate specimen
- Prepare a cartridge and run the Xpert® GI Panel
- Explain the Xpert® GI Panel control strategy
- Interpret the patient results report



# Training Agenda

- 1 [Overview](#)
- 2 [Kit Handling](#)
- 3 [Specimen Collection, Transport and Storage](#)
- 4 [Cartridge Preparation](#)
- 5 [Quality Controls](#)
- 6 [Results Interpretation](#)
- 7 [Troubleshooting](#)



# Overview



# The Cepheid Solution



- Qualitative detection of **DNA** and **RNA** targets from multiple bacteria, parasites and/or virus directly from stool in Cary Blair transport media
- On-board internal controls for each specimen
  - Probe Check Control (PCC)
  - Sample Processing Control (SPC)
  - Internal Control (IC)
- Results in approximately **74 minutes**
- Closed cartridge system minimizes risk of contamination
- On-demand results
- Random access





# Intended Use

- The Xpert® GI Panel, performed on the GeneXpert® Instrument Systems, is a qualitative multiplexed *in vitro* diagnostic test that is capable of the simultaneous detection and identification of DNA and RNA from multiple bacteria, parasites and/or virus directly from stool samples in Cary Blair transport media obtained from individuals with signs and symptoms of gastrointestinal infection. The test utilizes automated, qualitative real time polymerase chain reaction (PCR).





# Intended Use (Continued)

- The following bacteria (including several diarrheagenic *E. coli*/*Shigella* pathotypes), parasites, and virus are identified using the Xpert® GI Panel:

	Pathogens Detected	Pathogens Reported
<b>Bacteria</b>	<i>Campylobacter (C. jejuni/C. coli)</i>	<i>Campylobacter</i>
	Shiga toxin-producing <i>Escherichia coli</i> (STEC) <i>stx1/stx2</i>	STEC <i>stx1</i>
		STEC <i>stx2</i>
	<i>Salmonella</i>	<i>Salmonella</i>
	<i>Shigella</i> /Enteroinvasive <i>Escherichia coli</i>	<i>Shigella</i> EIEC
	<i>Yersinia enterocolitica</i>	<i>Yersinia</i>
	<i>Vibrio parahaemolyticus</i>	<i>V. parahaemolyticus</i>
	<i>Vibrio cholerae</i>	<i>V. cholerae</i>
<b>Parasites</b>	<i>Giardia</i> (also known as <i>G. intestinalis</i> , <i>G. duodenalis</i> & <i>G. lamblia</i> )	<i>Giardia</i>
	<i>Cryptosporidium</i>	<i>Cryptosporidium</i>
<b>Virus</b>	Norovirus GI/GII	Norovirus





# Intended Use (Continued)

- Results are meant to be used in conjunction with other clinical, laboratory and epidemiological data and should not be used as the sole basis for diagnosis, treatment or other patient management decisions. Positive results do not rule out co-infection with pathogens not included in the Xpert® GI Panel. The pathogen detected may not be the definite cause of the disease. Negative results in the setting of clinical illness compatible with gastroenteritis may be due to infection by pathogens that are not detected by this test or non-infectious causes such as ulcerative colitis, irritable bowel syndrome, or Crohn's disease.

## Intended User/Environment

The Xpert® GI Panel is intended to be performed by healthcare professionals trained on the use of the test. This test is for use in a laboratory environment.



# Xpert® GI Panel Requirements



## GeneXpert® Software

- GeneXpert® Dx software **v6.6** or higher
- Cepheid OS software **v2.3** or higher
- Xpertise software **v7.2** or higher
- GeneXpert® system equipped with 10-color modules

## Test Kits

- GXGI-CE-10

## Specimen Collection

- Stool in Cary Blair media

## Materials Required but Not Provided

- Cary Blair media

## Other Materials

- Personal Protective Equipment (PPE)
- 1:10 dilution bleach
- 70% ethanol or denatured ethanol
- Printer if needed



# Kit Handling





# Good Laboratory Practice Review

## Personal Protective Equipment (PPE)

- Wear clean lab coats, safety glasses, and gloves
- Change gloves between processing specimens

## Lab Bench Area

- Clean work surfaces routinely with:
  - ✓ 1:10 dilution of household bleach\*
  - ✓ 70% ethanol solution
- After cleaning, ensure work surfaces are dry and gloves are changed



## Specimen and Kit Storage

- Store specimens away from kit to prevent contamination

## Equipment

- Follow the manufacturer's requirements for instrument maintenance
- Setup specimens away from the instrument
- Use filtered pipette tips when recommended

\*Final Active Chlorine concentration should be 0.5% regardless of the household bleach concentration



# Xpert® GI Panel Kit Components



Catalog Number	GXGI-CE-10
Cartridges Per Kit	10
Disposable Transfer Swabs	10
CD	<p>CD (not pictured) contains the following:</p> <ul style="list-style-type: none"> <li>Assay Definition File (ADF)</li> <li>Instructions to import ADF into GeneXpert® software</li> <li>Instructions for use</li> </ul>
Storage	2-28° C



**➔ Cartridges contain chemically hazardous substances. Please see Instructions for Use and Safety Data Sheet for more detailed information.**



# Xpert® GI Panel Storage and Handling

- Store the Xpert® GI Panel cartridges at 2–28 °C until the expiration date provided on the label.
- If refrigerated, it is recommended to equilibrate the cartridge to room temperature (15-25 °C) before use.
- Do not open a cartridge until you are ready to perform testing.
- Do not use cartridges that have passed the expiration date.
- Do not use a cartridge that has leaked.



# Warnings and Precautions

- Do not use a cartridge that:
  - appears wet, has leaked or if the lid seal appears to have been broken
  - appears damaged
  - has been dropped after removing it from packaging
  - has been dropped or shaken after adding the specimen to it
  - has a damaged reaction tube
  - has been used; each cartridge is single-use to process one test
  - has expired
- Do not reuse transfer pipettes
- Specimens must be collected and tested before the expiration date of the Cary Blair Medium.
- Treat all biological specimens, including used cartridges, as if capable of transmitting infectious agents. Because it is often impossible to know which might be infectious, all biological specimens should be treated with standard precautions.





# Warnings and Precautions (Continued)

- Guidelines for specimen handling are available from the U.S. Centers for Disease Control and Prevention <sup>1</sup> and the Clinical and Laboratory Standards Institute.<sup>2</sup>
- Maintain proper storage conditions during specimen transport to ensure the integrity of the specimen (see Specimen Collection, Transport, and Storage Section). Specimen stability under shipping conditions other than those recommended has not been evaluated.
- In the event of specimens spilling, wear gloves and absorb the spill with paper towels. In the case of known or suspected *Cryptosporidium* or *Giardia*-containing samples, cover the spill area with paper towels and flood with 3% hydrogen peroxide. Allow a minimum of 20 minutes of contact time. Wipe the area dry <sup>3</sup>. If *Cryptosporidium* or *Giardia*-containing samples are not suspected, thoroughly clean the contaminated area with a 1:10 dilution of freshly prepared household chlorine bleach. Final active chlorine concentration should be 0.5% regardless of the household bleach concentration in your country. Allow a minimum of five minutes of contact time. Ensure the work area is dry before using 70% denatured ethanol to remove bleach residue. Allow surface to dry completely before proceeding. Or, follow your institution's standard procedures for a contamination or spill event. For equipment, follow the manufacturer's recommendations for decontamination of equipment.

1. Centers for Disease Control and Prevention. Biosafety in microbiological and biomedical laboratories. (refer to latest edition).

2. CLSI Publication M29. Protection of laboratory workers from occupationally acquired infections; Approved Guideline. (refer to latest edition).

3. [https://www.cdc.gov/parasites/crypto/resources/childcare\\_outbreak.pdf](https://www.cdc.gov/parasites/crypto/resources/childcare_outbreak.pdf)



# Specimen Collection, Transport and Storage





# Specimen Collection

Specimens should be transferred into **Cary Blair** media following the Cary Blair manufacturer's instructions or institutional guidelines. Specimens should be tested with the Xpert® GI Panel as soon as possible.



# Specimen Transport and Storage



## Specimen Type

Cary Blair medium

## Transport and Storage

15-25° C for up to 24 hours

2-8° C up to 4 days



Proper specimen collection, storage, and transport are critical to the performance of this test. Specimen stability under shipping and storage conditions other than those listed above have not been evaluated with the Xpert® GI Panel test.



# Cartridge Preparation





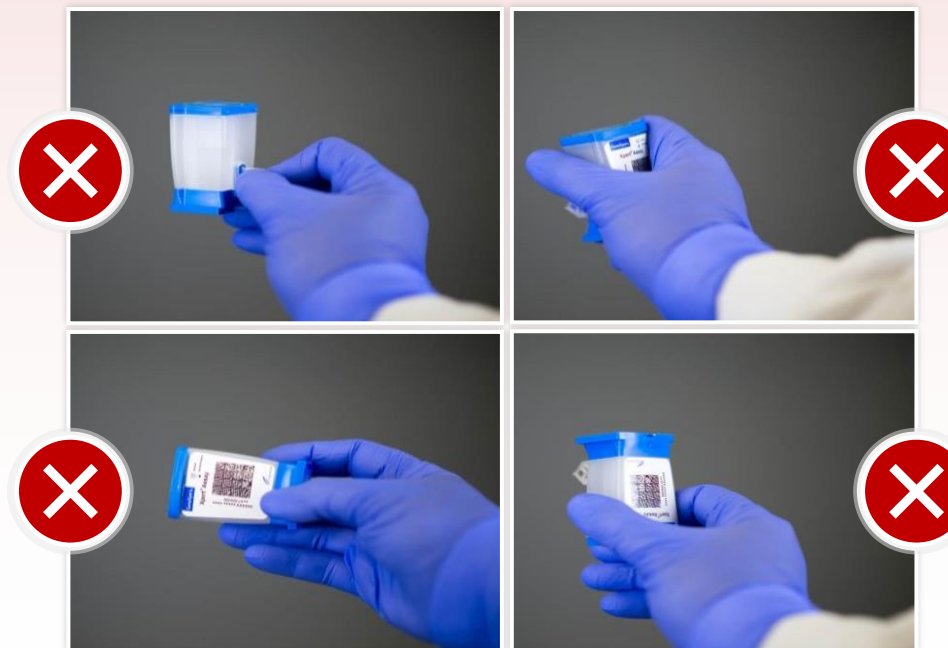
# Proper Cartridge Handling Techniques

## Correct



- Do not touch the reaction tube
- Keep the cartridge upright after seal has been broken
- Do not tilt when scanning the cartridge

## Incorrect




# Xpert® GI Panel Cartridge Preparation

## Xpert® GI Panel Cartridge Preparation

Refer to the Instructions for Use for detailed instructions, precautions, and warnings.

**1** Remove a cartridge from the package (it is recommended to equilibrate the cartridge to room temperature [15-25°C] before use).



**2** Mix specimen thoroughly until fully homogenized. Vortex can be used.




**3** Open the cartridge by lifting the lid.



**4** Remove the transfer swab from the wrapper.




**5** Dip the transfer swab for approximately 5 seconds into the sample and place the inoculated swab into the sample chamber of the Xpert GI Panel cartridge.



**6** Break the swab (at the break point) by snapping the shaft against the notch in the sample chamber opening leaving the swab tip in the sample chamber.

**NOTE:** There should not be any piece of the swab sticking out of the sample chamber.



**7** Close the cartridge lid.



# Which GeneXpert® System Do You Have?

## 1 GeneXpert Dx



## 2 GeneXpert System with Touchscreen



## 3 GeneXpert Infinity



For complete details on how to run a test, refer to the Instructions for Use and the system Operator Manual.

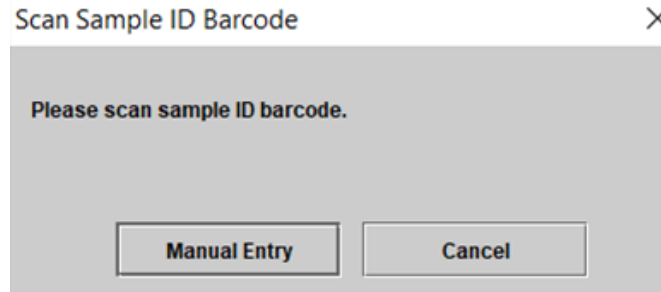
# Run a Test on GeneXpert® Dx

1 Start a test.



Start the test within **30 minutes** after adding the specimen to the cartridge.

2 Scan barcode for Patient and/or Sample ID.



3 Scan the cartridge.

To Scan do not click on Manual Entry or Cancel.





# Run a Test on GeneXpert® Dx (Continued)

4 Complete the fields as required.

5 Xpert® GI Panel test is selected automatically.

6 The module is selected automatically.

7 Click on Start Test.

8 A green light will flash on the module.  
Load the cartridge into module and close the door.

Create Test

Patient ID

Sample ID

Patient ID 2

Last Name

Name

Select Assay

Select Module

Reagent Lot ID\*  Expiration Date\*

Test Type

Sample Type  Other S

Notes



# Run a Test on GeneXpert® System with Touchscreen

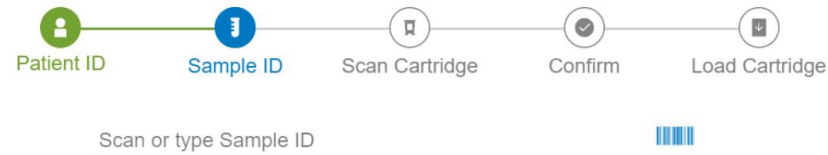


1 Start a test.

Start Test

Start the test within **30 minutes** after adding the specimen to the cartridge.

2 Scan barcode for Patient and/or Sample ID.



3 Scan the cartridge.



# Run a Test on GeneXpert® System with Touchscreen (Continued)



4 Click Confirm.

5 A green light will flash on the module. Load the cartridge into module and close the door.

The screenshot displays a confirmation screen on the GeneXpert touchscreen. At the top, a progress bar shows five steps: Patient ID, Sample ID, Scan Cartridge, Confirm (highlighted in blue), and Load Cartridge. Below the progress bar, there are several input fields and their corresponding values:

Patient ID	Cartridge S/N	251162261
Patient ID 2	Reagent Lot ID	05464
Last Name	Cartridge Exp. Date	04/08/29
First Name	Test Name	Xpert GI Panel
Sample ID		2023.02.23_15.24

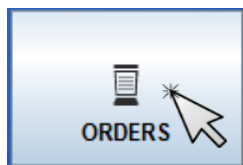
At the bottom of the screen, there are three buttons: 'Cancel' (grey), 'Back' (light blue), and 'Confirm' (dark blue). A status bar at the very bottom shows the date and time '02/23/23 15:24:30' and the user 'User: genexpert'.





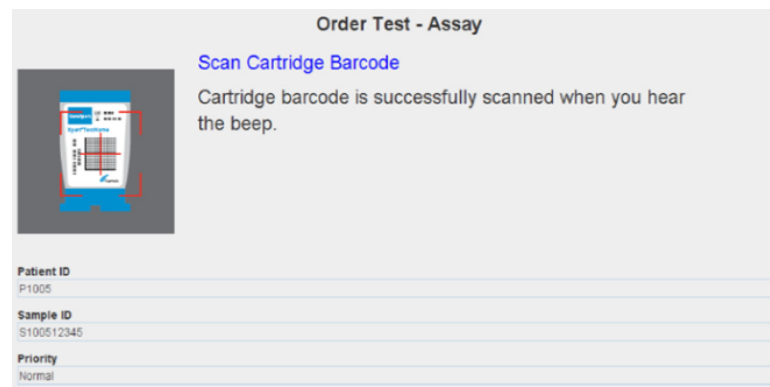
# Run a Test on GeneXpert<sup>®</sup> Infinity

1 Start a test.



Place the cartridge on the conveyor within **30 minutes** of adding the specimen.

2 Scan barcode for Patient and/or Sample ID.



3 Scan the cartridge.

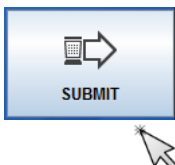


# Run a Test on GeneXpert® Infinity (Continued)

4 Complete the fields as required.

5 Xpert® GI Panel test is selected automatically.

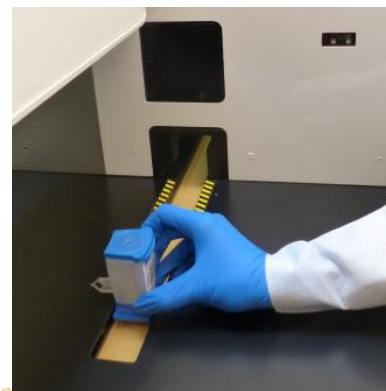
6 Click SUBMIT.



7 Place the cartridge on the conveyor belt.

Order Test - Test Information

<b>Patient ID</b> patientid	
<b>Sample ID</b> sampleid	
<b>Last Name</b> patient	<b>First Name</b> id
<b>Assay*</b> Xpert GI Panel	
<b>Reagent Lot ID*</b> 12102	<b>Cartridge S/N*</b> 282769448
<b>Expiration Date*</b> 2018/11/04	<b>Priority</b> Normal
<b>Test Type</b> Specimen	
<b>Sample Type</b> Other	<b>Other Sample Type</b>
<b>Notes</b>	





# Waste Disposal Warnings and Precautions

**Biological specimens, transfer devices, and used cartridges should be considered capable of transmitting infectious agents and require use of standard precautions.**

Follow your institution's environmental waste procedures for proper disposal of used cartridges and unused reagents.

.....  
These materials may exhibit characteristics of chemical hazardous waste requiring specific disposal.

.....  
**Please Note:** Used cartridges may contain potentially infectious materials, as well as highly amplified PCR target(s). Do not open or attempt to alter any part of the cartridge for disposal.

.....  
If country or regional regulations do not provide clear direction on proper disposal, biological specimens and used cartridges should be disposed per WHO [World Health Organization] medical waste handling and disposal guidelines.



# Quality Controls



# Xpert® GI Panel Control Strategy

## Xpert® GI Panel Quality Controls

- Each Xpert® cartridge is a self-contained test device
- Cepheid designed specific molecular methods to include internal controls that enable the system to detect specific failure modes within each cartridge:
  - Probe Check Control (PCC)
  - Sample Processing Control (SPC)
  - Internal Control (IC)



Refer to 301-4868 GeneXpert® Quality Control Features for all Cepheid Xpert® tests.



# Internal Quality Controls

## Probe Check Controls (PCC)

Before the PCR step, fluorescence signal is measured on all probes and compared with default factory settings to monitor:

- Bead rehydration
- Reaction tube filling
- Probe integrity
- Dye stability

## Sample Processing Controls (SPC)

The SPC is present to control for adequate processing of the specimen and to monitor for the presence of potential inhibitor(s) in the PCR reaction.

- Verifies the lysis of hard to lyse pathogens (parasites and bacteria)
- Verifies adequate extraction and amplification of the specimen
- Detects PCR inhibition
- Ensures appropriate PCR conditions for amplification (temperature and time)
- Verifies that PCR reagents are functional
- Acts as a control for functionality of melt curve analysis
- Must be positive in a negative specimen to be a valid test
- Can be positive or negative in a positive specimen



# Internal Quality Controls (*Continued*)

## Internal Control (IC)

The IC is an RNA control included in each cartridge to verify release of RNA from the sample.

- Verifies the sample processing is adequate.
- Detects specimen-associated inhibition of the reverse transcription and PCR reactions.
- Must be positive in a negative sample. Can be negative or positive in a positive sample.



# Commercially Available External Controls

## Microbiologics® Gastrointestinal Control Panel (Inactivated Swab)

Contents	Part Number	Total Vials/Boxes
<b>Positive Control</b>	8248	6 individually packaged lyophilized positive control swabs
<b>Negative Control</b>	8248	6 individually packaged lyophilized negative control swabs

**NOTE:** To minimize degradation of the control material, return unopened QC to the manufacturer's recommended storage conditions immediately after use.



# Commercially Available External Controls (Continued)

The external quality control materials provided by the specified vendor's IFU are an optional source. All external controls must be used in accordance with local, state, and/or federal regulations or accreditation requirements, as applicable.



# Results Interpretation





# Xpert® GI Panel Targets and Results

Target	Result	
<i>Campylobacter</i>	NEGATIVE	POSITIVE
<i>Salmonella</i>		
STEC stx1		
STEC stx2		
<i>Shigella EIEC</i>		
<i>V. cholerae</i>		
<i>V. parahaemolyticus</i>		
<i>Yersinia</i>		
<i>Cryptosporidium</i>		
<i>Giardia</i>		
Norovirus		

The results are interpreted automatically by the GeneXpert® system software and shown in the **View Results** window. The test provides test results based on the detection of respective gene targets according to the algorithms in the ADF.



# Which GeneXpert® System Do You Have?

## 1 GeneXpert Dx



## 2 GeneXpert System with Touchscreen



## 3 GeneXpert Infinity



For complete details on how to run a test, refer to the Instructions for Use and the system Operator Manual.

# GeneXpert® Dx GI Panel POSITIVE

## Example Report

GeneXpert PC Test Report

Patient ID: Positive QC for Ct  
Sample ID: Specimen  
Test Type:  
Sample Type:

Assay Information

Assay	Assay Version	Assay Type
Xpert GI Panel	1	In Vitro Diagnostic

Test Result:

- Campylobacter NEGATIVE;
- Salmonella POSITIVE;
- STEC stx1 NEGATIVE;
- STEC stx2 NEGATIVE;
- Shigella EIEC POSITIVE;
- V. cholerae NEGATIVE;
- V. parahaemolyticus NEGATIVE;
- Yersinia NEGATIVE;
- Cryptosporidium POSITIVE;
- Giardia POSITIVE;
- Norovirus POSITIVE

Analyte Result

Analyte Name	Ct	EndPt	Analyte Result	Probe Check Result
SPC	32.7	174	POS	PASS
IC	28.4	328	POS	PASS
Salmonella	23.4	306	POS	PASS
Shigella	20.1	425	POS	PASS
EIEC				
Cryptosporidium	13.2	422	POS	PASS
Giardia	16.2	477	POS	PASS
Norovirus	15.7	453	POS	PASS

GeneXpert PC Test Report

Melt Peaks

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	71.7	11.2
Campylobacter		
STEC stx1		
STEC stx2		
V. cholerae		
V. parahaemolyticus		
Yersinia		

- The *Salmonella*, *Shigella EIEC*, *Cryptosporidium*, *Giardia*, and/or *Norovirus* signal has a Ct within the valid cycle range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) are detected.
- IC – NA; IC is ignored because Target(s) are detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**



# GeneXpert® Dx GI Panel POSITIVE

## Example Report

GeneXpert PC Test Report

Patient ID: Positive QC for Melts  
 Sample ID: Specimen  
 Test Type: Specimen  
 Sample Type: Specimen

Assay Information

Assay	Assay Version	Assay Type
Xpert GI Panel	1	In Vitro Diagnostic

Test Result:

Campylobacter POSITIVE;  
 Salmonella NEGATIVE;  
 STEC stx1 POSITIVE;  
 STEC stx2 POSITIVE;  
 Shigella EIEC NEGATIVE;  
 V. cholerae POSITIVE;  
 V. parahaemolyticus POSITIVE;  
 Yersinia POSITIVE;  
 Cryptosporidium NEGATIVE;  
 Giardia NEGATIVE;  
 Norovirus NEGATIVE

Analyte Result

Analyte Name	Ct	EndPt	Analyte Result	Probe Check Result
SPC	30.4	194	POS	PASS
IC	28.8	330	POS	PASS
Salmonella	0.0	1	NEG	PASS
Shigella EIEC	0.0	-1	NEG	PASS
Cryptosporidium	0.0	8	NEG	PASS
um	0.0	1	NEG	PASS
Giardia	0.0	1	NEG	PASS
Norovirus	0.0	1	NEG	PASS

GeneXpert PC Test Report

Melt Peaks

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	71.7	11.2
Campylobacter	60.6	28.2
STEC stx1	74.9	49.5
STEC stx2	76.9	20.5
V. cholerae	61.1	20.6
V. parahaemolyticus	66.2	20.9
Yersinia	61.8	21.5

- The *Campylobacter*, STEC stx1, STEC stx2, *V. cholerae*, *V. parahaemolyticus*, and/or *Yersinia* signal has a melt peak above the target specific threshold setting and within the melt temperature (Tm) valid range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) detected.
- IC – NA; IC is ignored because Target(s) detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**

Melt Peak data is not included on the test report when run on the GeneXpert® Dx software. If needed, the Melt Peaks data can be added to the Dx test report by reprinting it and selecting the Melt Peaks box. The Melt Peaks can also be accessed in the Melt Peaks tab in the GeneXpert® Dx software.



# GeneXpert® Dx GI Panel NEGATIVE



## Example Report

GeneXpert PC Test Report 10/14/25 15:30:21

Patient ID:  
Sample ID: Positive QC for Melts  
Test Type: Specimen  
Sample Type:

Assay Information

Assay	Assay Version	Assay Type
Xpert GI Panel	1	In Vitro Diagnostic

Test Result:

Campylobacter NEGATIVE;  
Salmonella NEGATIVE;  
STEC stx1 NEGATIVE;  
STEC stx2 NEGATIVE;  
Shigella EIEC NEGATIVE;  
V. cholerae NEGATIVE;  
V. parahaemolyticus NEGATIVE;  
Yersinia NEGATIVE;  
Cryptosporidium NEGATIVE;  
Giardia NEGATIVE;  
Norovirus NEGATIVE

Analyte Result

Analyte Name	Ct	EndPt	Analyte Result	Probe Check Result
SPC	30.4	194	POS	PASS
IC	28.8	330	POS	PASS
Salmonella	0.0	1	NEG	PASS
Shigella EIEC	0.0	-1	NEG	PASS
Cryptosporidium	0.0	8	NEG	PASS
Giardia	0.0	1	NEG	PASS
Norovirus	0.0	1	NEG	PASS

GeneXpert PC Test Report 10/14/25 15:30:21

Melt Peaks

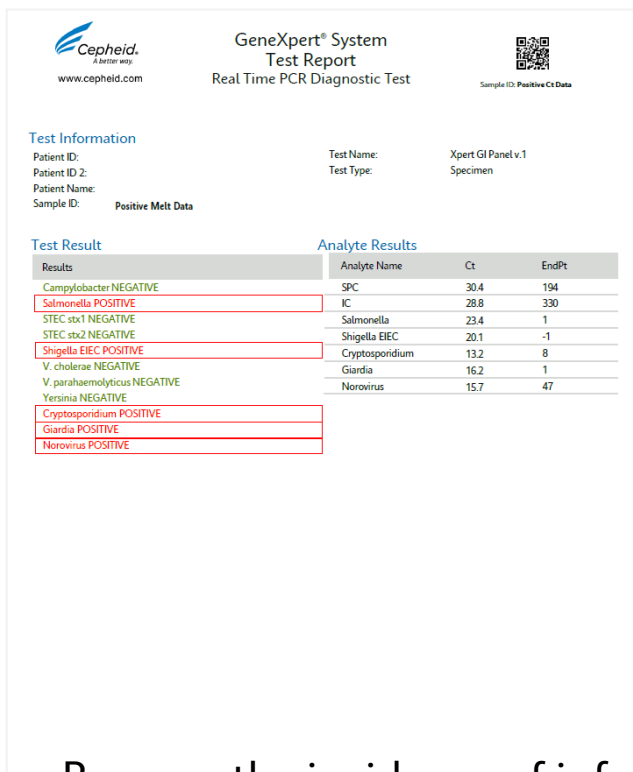
Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	71.7	11.2
Campylobacter	60.6	28.2
STEC stx1	74.9	49.5
STEC stx2	76.9	20.5
V. cholerae	61.1	20.6
V. parahaemolyticus	66.2	20.9
Yersinia	61.8	21.5

- None of the pathogens listed are detected.
- SPC - PASS
  - The SPC signal has a Ct within the valid cycle range AND
  - The SPC signal has a melt peak above the target specific threshold setting and within the melt temperature (Tm) valid range.
- IC – PASS
  - The IC signal has a Ct within the valid cycle range.
- Probe Check - PASS; all probe check results pass.



# GeneXpert® System with Touchscreen GI Panel POSITIVE

## Example Report



**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: Positive Ct Data

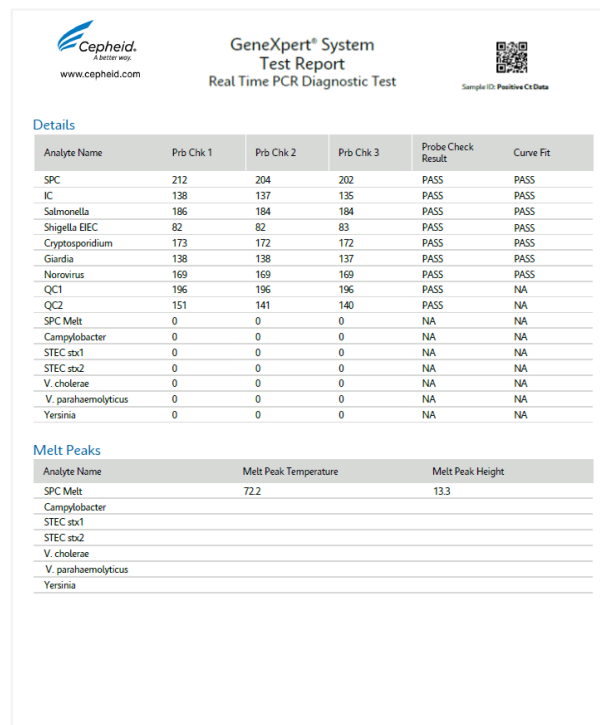
**Test Information**

Patient ID: \_\_\_\_\_  
Patient ID 2: \_\_\_\_\_  
Patient Name: \_\_\_\_\_  
Sample ID: Positive Melt Data

Test Name: Xpert GI Panel v.1  
Test Type: Specimen

**Test Result**

Results	Analyte Name	Ct	EndPt
Campylobacter NEGATIVE	SPC	30.4	194
Salmonella POSITIVE	IC	28.8	330
STEC stx1 NEGATIVE	Salmonella	23.4	1
STEC stx2 NEGATIVE	Shigella EIEC	20.1	-1
Shigella EIEC POSITIVE	Cryptosporidium	13.2	8
V. cholerae NEGATIVE	Giardia	16.2	1
V. parahaemolyticus NEGATIVE	Norovirus	15.7	47
Yersinia NEGATIVE			
Cryptosporidium POSITIVE			
Giardia POSITIVE			
Norovirus POSITIVE			



**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: Positive Ct Data

**Details**

Analyte Name	Prb Chk 1	Prb Chk 2	Prb Chk 3	Probe Check Result	Curve Fit
SPC	212	204	202	PASS	PASS
IC	138	137	135	PASS	PASS
Salmonella	196	184	184	PASS	PASS
Shigella EIEC	82	82	83	PASS	PASS
Cryptosporidium	173	172	172	PASS	PASS
Giardia	138	138	137	PASS	PASS
Norovirus	169	169	169	PASS	PASS
QC1	196	196	196	PASS	NA
QC2	151	141	140	PASS	NA
SPC Melt	0	0	0	NA	NA
Campylobacter	0	0	0	NA	NA
STEC stx1	0	0	0	NA	NA
STEC stx2	0	0	0	NA	NA
V. cholerae	0	0	0	NA	NA
V. parahaemolyticus	0	0	0	NA	NA
Yersinia	0	0	0	NA	NA

**Melt Peaks**

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	72.2	13.3
Campylobacter		
STEC stx1		
STEC stx2		
V. cholerae		
V. parahaemolyticus		
Yersinia		

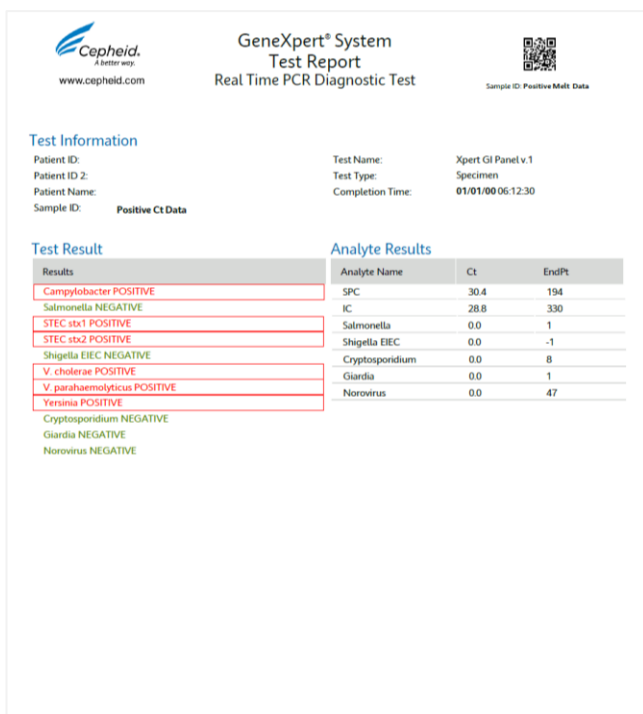
- The *Salmonella*, *Shigella EIEC*, *Cryptosporidium*, *Giardia*, and/or *Norovirus* signal has a Ct within the valid cycle range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) are detected.
- IC – NA; IC is ignored because Target(s) are detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**



# GeneXpert® System with Touchscreen GI Panel POSITIVE

## Example Report



**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: Positive Melt Data

**Test Information**

Patient ID: \_\_\_\_\_  
 Patient ID 2: \_\_\_\_\_  
 Patient Name: \_\_\_\_\_  
 Sample ID: Positive Ct Data

Test Name: Xpert GI Panel v.1  
 Test Type: Specimen  
 Completion Time: 01/01/00 06:12:30

**Test Result**

Results
Campylobacter POSITIVE
Salmonella NEGATIVE
STEC stx1 POSITIVE
STEC stx2 POSITIVE
Shigella EIEC NEGATIVE
V. cholerae POSITIVE
V. parahaemolyticus POSITIVE
Yersinia POSITIVE
Cryptosporidium NEGATIVE
Giardia NEGATIVE
Norovirus NEGATIVE

**Analyte Results**

Analyte Name	Ct	EndPt
SPC	30.4	194
IC	28.8	330
Salmonella	0.0	1
Shigella EIEC	0.0	-1
Cryptosporidium	0.0	8
Giardia	0.0	1
Norovirus	0.0	47



**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: Positive Melt Data

**Details**

Analyte Name	Prb Chk 1	Prb Chk 2	Prb Chk 3	Probe Check Result	Curve Fit
SPC	193	181	177	PASS	PASS
IC	183	183	181	PASS	PASS
Salmonella	203	201	200	PASS	PASS
Shigella EIEC	83	82	83	PASS	PASS
Cryptosporidium	170	165	163	PASS	PASS
Giardia	114	113	112	PASS	PASS
Norovirus	144	143	141	PASS	PASS
QC1	189	189	188	PASS	NA
QC2	142	128	126	PASS	NA
SPC Melt	0	0	0	NA	NA
Campylobacter	0	0	0	NA	NA
STEC stx1	0	0	0	NA	NA
STEC stx2	0	0	0	NA	NA
V. cholerae	0	0	0	NA	NA
V. parahaemolyticus	0	0	0	NA	NA
Yersinia	0	0	0	NA	NA

**Melt Peaks**

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	71.7	11.2
Campylobacter	60.6	28.2
STEC stx1	74.9	49.5
STEC stx2	76.9	20.5
V. cholerae	61.1	20.6
V. parahaemolyticus	66.2	20.9
Yersinia	61.8	21.5

- The *Campylobacter*, STEC stx1, STEC stx2, *V. cholerae*, *V. parahaemolyticus*, and/or *Yersinia* signal has a melt peak above the target specific threshold setting and within the melt temperature (T<sub>m</sub>) valid range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) detected.
- IC – NA; IC is ignored because Target(s) detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**



# GeneXpert® System with Touchscreen GI Panel NEGATIVE



## Example Report

www.cephheid.com

**GeneXpert® System  
Test Report**  
Real Time PCR Diagnostic Test

Sample ID: LIS\_US\_IVD\_Negative

www.cephheid.com

**Test Information**

Patient ID: \_\_\_\_\_  
 Patient ID 2: \_\_\_\_\_  
 Patient Name: \_\_\_\_\_  
 Sample ID: LIS\_US\_IVD\_Negative

Test Name: Xpert GI Panel v.1  
 Test Type: Specimen

**Test Result**

Results
Campylobacter NEGATIVE
Salmonella NEGATIVE
STEC stx1 NEGATIVE
STEC stx2 NEGATIVE
Shigella EIEC NEGATIVE
V. cholerae NEGATIVE
V. parahaemolyticus NEGATIVE
Yersinia NEGATIVE
Cryptosporidium NEGATIVE
Giardia NEGATIVE
Norovirus NEGATIVE

**Analyte Results**

Analyte Name	Ct	EndPt
SPC	30.4	194
IC	28.8	330
Salmonella	0.0	1
Shigella EIEC	0.0	-1
Cryptosporidium	0.0	8
Giardia	0.0	1
Norovirus	48.7	47

**Details**

Analyte Name	Prb Chk 1	Prb Chk 2	Prb Chk 3	Probe Check Result	Curve Fit
SPC	212	204	202	PASS	PASS
IC	138	137	135	PASS	PASS
Salmonella	186	184	184	PASS	NA
Shigella EIEC	82	82	83	PASS	NA
Cryptosporidium	173	172	172	PASS	NA
Giardia	138	138	137	PASS	NA
Norovirus	169	169	169	PASS	NA
QC1	196	196	196	PASS	NA
QC2	151	141	140	PASS	NA
SPC Melt	0	0	0	NA	NA
Campylobacter	0	0	0	NA	NA
STEC stx1	0	0	0	NA	NA
STEC stx2	0	0	0	NA	NA
V. cholerae	0	0	0	NA	NA
V. parahaemolyticus	0	0	0	NA	NA
Yersinia	0	0	0	NA	NA

**Melt Peaks**

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	72.2	13.3
Campylobacter		
STEC stx1		
STEC stx2		
V. cholerae		
V. parahaemolyticus		
Yersinia		

- None of the pathogens listed are detected.
- SPC - PASS
  - The SPC signal has a Ct within the valid cycle range AND
  - The SPC signal has a melt peak above the target specific threshold setting and within the melt temperature (Tm) valid range.
- IC – PASS
  - The IC signal has a Ct within the valid cycle range.
- Probe Check - PASS; all probe check results pass.



# GeneXpert® Infinity GI Panel POSITIVE

## Example Report



GeneXpert® System Test Report		Real Time PCR Diagnostic Test																																																																							
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GeneXpert® System Test Report		Real Time PCR Diagnostic Test																									
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- The *Salmonella*, *Shigella EIEC*, *Cryptosporidium*, *Giardia*, and/or *Norovirus* signal has a Ct within the valid cycle range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) are detected.
- IC – NA; IC is ignored because Target(s) are detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**



# GeneXpert® Infinity GI Panel POSITIVE

## Example Report



GeneXpert® System Test Report						
Real Time PCR Diagnostic Test						
Sample ID: Positive QC for Melt Peaks						
<b>Test Information</b>						
Patient ID:	Test Type: Specimen					
Sample ID: Positive QC for Melt Peaks						
Test Name: Xpert GI Panel v.1						
<b>Test Result:</b>						
<b>Results</b>	<b>Analyte Results</b>					
Campylobacter POSITIVE	SPC 30.4					
Salmonella NEGATIVE	IC 28.8					
STEC stx1 POSITIVE	Salmonella 0.0					
STEC stx2 POSITIVE	Shigella EIEC 0.0					
Shigella EIEC NEGATIVE	Cryptosporidium 0.0					
V. cholerae POSITIVE	Giardia 0.0					
V. parahaemolyticus POSITIVE	Norovirus 0.0					
Yersinia POSITIVE						
Cryptosporidium NEGATIVE						
Giardia NEGATIVE						
Norovirus NEGATIVE						
<b>Details</b>						
Analyte Name	Prb Chk 1	Prb Chk 2	Prb Chk 3	Probe Check Result	2nd Deriv Peak Height	Curve Fit
SPC	212	204	202	PASS	0.0	PASS
IC	138	137	135	PASS	0.0	PASS
Salmonella	186	184	184	PASS	0.0	NA
Shigella EIEC	82	82	83	PASS	0.0	NA
Cryptosporidium	173	172	172	PASS	0.0	NA
Giardia	138	138	137	PASS	0.0	NA
Norovirus	169	169	169	PASS	0.0	NA
QC1	196	196	196	PASS	0.0	NA

GeneXpert® System Test Report		
Real Time PCR Diagnostic Test		
Sample ID: Positive QC for Melt Peaks		
<b>Melt Peaks</b>		
Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	71.7	11.2
Campylobacter	60.6	28.2
STEC stx1	74.9	49.5
STEC stx2	76.9	20.5
V. cholerae	61.1	20.6
V. parahaemolyticus	66.2	20.9
Yersinia	61.8	21.5

- The *Campylobacter*, STEC stx1, STEC stx2, *V. cholerae*, *V. parahaemolyticus*, and/or *Yersinia* signal has a melt peak above the target specific threshold setting and within the melt temperature (Tm) valid range.
- Except Target(s) detected, all other pathogens are not detected.
- Probe Check: PASS; all probe check results pass.
- SPC – Not Applicable (NA); SPC is ignored because Target(s) detected.
- IC – NA; IC is ignored because Target(s) detected.

Because the incidence of infection with multiple (3 or more) agents are low, it is recommended that specimens undergo repeat testing if nucleic acids from multiple analytes are detected in a single specimen. **This does not apply when running Quality Control.**



# GeneXpert® Infinity GI Panel NEGATIVE

## Example Report



**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: LIS\_US IVD\_Negative

**Test Information**

Patient ID: \_\_\_\_\_ Test Type: Specimen  
 Sample ID: LIS\_US IVD\_Negative  
 Test Name: Xpert GI Panel v.1

**Test Result:**

Results	Analyte Name	Ct
Campylobacter NEGATIVE	SPC	30.4
Salmonella NEGATIVE	IC	28.8
STEC stx1 NEGATIVE	Salmonella	0.0
STEC stx2 NEGATIVE	Shigella EIEC	0.0
Shigella EIEC NEGATIVE	Cryptosporidium	0.0
V. cholerae NEGATIVE	Giardia	0.0
V. parahaemolyticus NEGATIVE	Norovirus	48.7
Yersinia NEGATIVE		
Cryptosporidium NEGATIVE		
Giardia NEGATIVE		
Norovirus NEGATIVE		

**Details**

Analyte Name	Prb Chk 1	Prb Chk 2	Prb Chk 3	Probe Check Result	2nd Deriv Peak Height	Curve Fit
SPC	212	204	202	PASS	0.0	PASS
IC	138	137	135	PASS	0.0	PASS
Salmonella	186	184	184	PASS	0.0	NA
Shigella EIEC	82	82	83	PASS	0.0	NA
Cryptosporidium	173	172	172	PASS	0.0	NA
Giardia	138	138	137	PASS	0.0	NA
Norovirus	169	169	169	PASS	0.0	NA
QC1	196	196	196	PASS	0.0	NA
QC2	151	141	140	PASS	0.0	NA

**GeneXpert® System Test Report**  
Real Time PCR Diagnostic Test

Sample ID: LIS\_US IVD\_Negative

SPC Melt	0	0	0	NA	0.0	NA
Campylobacter	0	0	0	NA	0.0	NA
STEC stx1	0	0	0	NA	0.0	NA
STEC stx2	0	0	0	NA	0.0	NA
V. cholerae	0	0	0	NA	0.0	NA
V. parahaemolyticus	0	0	0	NA	0.0	NA
Yersinia	0	0	0	NA	0.0	NA

**Melt Peaks**

Analyte Name	Melt Peak Temperature	Melt Peak Height
SPC Melt	72.2	13.3
Campylobacter		
STEC stx1		
STEC stx2		
V. cholerae		
V. parahaemolyticus		
Yersinia		

- None of the pathogens listed are detected.
- SPC - PASS
  - The SPC signal has a Ct within the valid cycle range AND
  - The SPC signal has a melt peak above the target specific threshold setting and within the melt temperature (Tm) valid range.
- IC – PASS
  - The IC signal has a Ct within the valid cycle range.
- Probe Check - PASS; all probe check results pass.



# Troubleshooting





# Reasons to Repeat the Test

## INVALID

Sample Processing Control (SPC) and/or Internal Control (IC) does not meet acceptance criteria and none of the target pathogens meet criteria for the test result POSITIVE.

Repeat test according to the Retest Procedure.

Probe Check: PASS; All probe check results pass

## ERROR

None of the target pathogens meet criteria for the test result POSITIVE or NEGATIVE.  
Repeat test according to the Retest Procedure.

SPC and IC: NO RESULT

Probe Check: FAIL; All or one of the probe check results failed.  
If the probe check passes, the error is caused by the maximum pressure limit exceeding the acceptable range, or by a system component failure.

## NO RESULT

Test result POSITIVE or NEGATIVE of the target pathogens cannot be determined.  
Repeat test according to the Retest Procedure.

Targets – NO RESULT

Probe Check: NA (not applicable)



# Retest Procedure

1



Discard used cartridge. Follow your institution's safety guidelines for disposal of cartridges.

2



If the result is **INVALID, ERROR, NO RESULT** or the result indicates infection with multiple pathogens, (3 or more) use a new cartridge to retest the specimen.

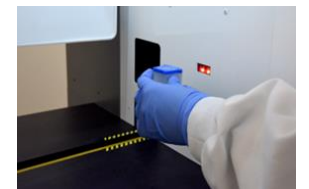
3



Obtain a new cartridge.

Process the specimen per the Instructions For Use.

4



Run the test on the system.



# Technical Assistance

- Before contacting Cepheid Technical Support, collect the following information:
  - Product name
  - Lot number
  - Serial number of the System
  - Software version and, if applicable, computer service tag number
  - Error messages (if any)
- Log your case online using the following link  
<https://my.cepheid.com/contact/technical-support>





*Thank You*

[www.Cepheid.com](http://www.Cepheid.com)

**End Show**

